

What we know about nonsteroidal anti-inflammatory drug hypersensitivity

Duy Le Pham^{1,2}, Ji-Hye Kim¹, Tu Hoang Kim Trinh¹, and Hae-Sim Park^{1,2}

¹Department of Allergy and Clinical Immunology, Ajou University School of Medicine, Suwon; ²Department of Biomedical Sciences, The Graduate School, Ajou University, Suwon, Korea

Received: February 14, 2016
Accepted: March 5, 2016

Correspondence to
Hae-Sim Park, M.D.

Department of Allergy and Clinical Immunology, Ajou University Hospital, 164 World cup-ro, Yeongtong-gu, Suwon 16499, Korea

Tel: +82-31-219-5150

Fax: +82-31-219-5154

E-mail: hspark@ajou.ac.kr

Nonsteroidal anti-inflammatory drugs (NSAIDs) are widely prescribed for the treatment of inflammatory diseases, but their use is frequently related to hypersensitivity reactions. This review outlines our current knowledge of NSAID hypersensitivity (NHS) with regard to its pathogenic, molecular, and genetic mechanisms, as well as diagnosis and treatment. The presentation of NHS varies from a local (skin and/or airways) reaction to systemic reactions, including anaphylaxis. At the molecular level, NHS reactions can be classified as cross-reactive (mediated by cyclooxygenase inhibition) or selective (specific activation of immunoglobulin E antibodies or T cells). Genetic polymorphisms and epigenetic factors have been shown to be closely associated with NHS, and may be useful as predictive markers. To diagnose NHS, inhalation or oral challenge tests are applied, with the exclusion of any cross-reactive NSAIDs. For patients diagnosed with NHS, absolute avoidance of NSAIDs/ aspirin is essential, and pharmacological treatment, including biologics, is often used to control their respiratory and cutaneous symptoms. Finally, desensitization is recommended only for selected patients with NHS. However, further research is required to develop new diagnostic methods and more effective treatments against NHS.

Keywords: Anti-inflammatory agents, non-steroidal; Drug hypersensitivity; Etiology; Genetic predisposition to disease

INTRODUCTION

Nonsteroidal anti-inflammatory drugs (NSAIDs) are widely prescribed for the treatment of various inflammatory diseases. However, NSAIDs can cause drug hypersensitivity reactions, varying from local reactions in the skin and/or airways to systemic symptoms, including life-threatening anaphylaxis [1,2]. NSAID hypersensitivity (NHS) has been reported in 4.3% to 11% of patients with asthma [3] and in 27% to 35% of patients with chronic urticaria (CU) [4]. Moreover, NHS contributes to 13.3% of anaphylaxis cases in Korean adults [5]. While various terms (e.g., sensitivity, idiosyncrasy, or intolerance) have been used to describe unintended and unpredictable adverse drug

reactions, the term “hypersensitivity” seems appropriate to describe the adverse reaction to NSAIDs, as its pathogenesis can involve both immunological and nonimmunological reactions [6,7].

NHS reactions may be classified as cross-reactive against several and chemically unrelated NSAIDs, or selective for a single compound or a group of chemically related compounds. Cross-reactive NHS reactions can manifest as three major clinical phenotypes: (1) NSAID-exacerbated respiratory disease (NERD), occurring in patients with underlying chronic upper and lower airway respiratory diseases; (2) NSAID-exacerbated cutaneous disease (NECD), occurring in patients with a history of CU; and (3) NSAID-induced urticaria/an-

gioedema (NIUA), occurring in otherwise healthy subjects (without a history of CU) [2,8]. Anaphylaxis is commonly seen in NIUA. In addition, cross-reactive NHS may induce both respiratory and cutaneous symptoms in a subgroup of patients [9]. Selective NHS reactions manifest as two clinical phenotypes: (1) single NIUA/anaphylaxis reactions, which develop immediately after drug intake (immunoglobulin E [IgE]-mediated); and (2) NSAID-induced delayed hypersensitivity reactions, which develop more than 24 hours after drug intake (T cell-mediated) [2].

The pathogenesis of NHS reactions is thought to be related to various mechanisms, including: (1) a non-immunological mechanism, through the inhibition of enzymes involved in the arachidonic acid (AA) pathway; and (2) an immunological mechanism, involving the production of drug-specific IgE antibodies or T cell activation. Moreover, genetic polymorphisms and epigenetic factors have been implicated in the pathogenesis of, and susceptibility to, NHS. This review summarizes current knowledge regarding the pathogenic mechanisms, associated genetic and epigenetic factors, diagnostic methods, and treatments for patients with NHS.

PATHOGENIC MECHANISMS OF NHS

The proposed pathogenic mechanisms and clinical manifestations of NHS are shown in Table 1 and Fig. 1 [10]. NHS reactions can be subdivided as cross-reactive, in which the cyclooxygenase 1 (COX-1) pathway is inhibited, or selective, resulting from an immunological (allergic)

reaction to the drugs and characterized by IgE-mediated (acute) or T cell-mediated (delayed) reactions [2,11-13]. In addition, novel mechanisms related to oxidative stress and platelet activation that could contribute to the pathogenesis of NHS have been reported [14-17].

Metabolism of arachidonic acid and the effects of NSAIDs

AA can be metabolized by either the COX pathway or the 5-lipoxygenase (5-LOX) pathway, with conversion to prostanoids or cysteinyl leukotrienes (CysLTs), respectively [18]. There are two isoforms of COX: COX-1 (expressed constitutively in most cells) and COX-2 (expressed either constitutively or in responses to inflammatory stimuli) [2]. The COX enzymes catalyze AA to prostaglandin (PG) G₂ and then to PGH₂, proceeding to the syntheses of PGE₂, PGI₂, PGD₂, PGF₂α, and thromboxane A₂ (TXA₂) by tissue-specific isomerase and synthase enzymes [18]. During prostanoid syntheses, COX-1 couples preferentially to thromboxane synthase (TBX-AS), PGF synthase, and cytosolic PGE synthase (PGES) isozymes, while COX-2 shows preferential coupling with PGI synthase and microsomal PGES isozymes [19]. In the 5-LOX pathway, 5-LOX converts AA into LTA₄, which is then converted to LTB₄ (by LTA₄ hydrolase) or catalyzed to LTC₄ (by LTC₄ synthase), followed by LTD₄ and then E₄ syntheses [20].

NSAIDs act by suppressing the activities of (1) COX isoenzymes, and (2) the enzymes responsible for prostanoid biosynthesis from AA [18,21,22]. Certain NSAIDs preferentially inhibit COX-1 and only partially inhibit COX-2 (e.g., aspirin, indomethacin, naproxen, and di-

Table 1. Pathogenic mechanisms and clinical manifestations of NSAID hypersensitivity

Type of reaction	Time of onset ^a	Clinical manifestation	Mechanism
Cross reactive	Acute	NERD	COX-1 inhibition
			Oxidative stress
		NECD	COX-1 inhibition
		NIUA	COX-1 inhibition
			Platelet activation
Selective	Acute	SNIUAA	IgE-mediated
	Delayed	NIDHR	T cell-mediated

NSAID, nonsteroidal anti-inflammatory drug; NERD, NSAID-exacerbated respiratory disease; COX-1, cyclooxygenase 1; NECD, NSAID-exacerbated cutaneous disease; NIUA, NSAID-induced urticaria/angioedema; SNIUAA, single-NSAID-induced urticaria/anaphylaxis; IgE, immunoglobulin E; NIDHR, NSAID-induced delayed hypersensitivity reaction.

^aAcute onset, clinical symptoms occur immediately or a few hours after drug administration; delayed onset, clinical symptoms occur 24 hours or more after drug administration.

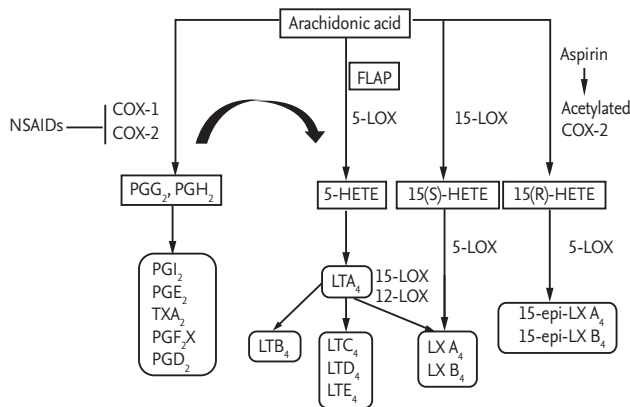


Figure 1. Arachidonic acid (AA) metabolism involved in the pathogenic mechanisms of nonsteroidal anti-inflammatory drug (NSAID) hypersensitivity. AA can be metabolized by either the cyclooxygenase (COX) pathway or 5-lipoxygenase (5-LOX) pathway [25]. Inhibition of COX enzymes by NSAIDs shifts the conversion of AA to the 5-LOX pathway, leading to decreased production of prostaglandins (PGs) and increased production of cysteinyl leukotrienes (LTs). Lipoxin (LX) A_4/B_4 are synthesized either from LTA_4 or from 15S-hydroxyeicosatetraenoic acid (15(S)-HETE). In addition, aspirin acetylates COX-2, transforming AA to 15(R)-HETE and then to 15-epi-LX by 5-LOX [125]. In NSAID-exacerbated respiratory disease patients, the production of LX and 15-epi-LX, which have anti-inflammatory effects, could be decreased, possibly due to the excessive usage of AA for LT synthesis. FLAP, 5-lipoxygenase-activating protein.

clofenac); thereby, inhibiting production of protective PGs. Newer NSAIDs that inhibit COX-2 primarily (e.g., nimesulide, meloxicam) or specifically (e.g., celecoxib, rofecoxib) can suppress the inflammatory prostanoids, and only slightly decrease protective PG production [2].

COX-1 inhibition

By suppressing COX-1 activity, aspirin and some classical NSAIDs inhibit the synthesis of PGs, especially PGE_2 ; thereby, increasing the production of CysLTs (LTA_4 , LTB_4 , LTC_4 , LTD_4) [2,17,23,24]. PGE_2 has a protective effect against bronchoconstriction, releases inflammatory mediators from mast cells, and recruits immune cells to inflammatory sites [25,26]. On the other hand, CysLTs induce bronchoconstriction, airway inflammation, cell recruitment, and platelet activation [27-30]. In addition, aspirin can acetylate COX-2, leading to the generation of 15-hydroeicosatetraenoic acid (15-HETE), which is then transformed into lipoxin (LX) or 15-epimer-LX (15-epi-LX) by 5-LOX. Both LX and 15-epi-LX exert anti-inflam-

matory effects by inhibiting neutrophil and eosinophil chemotaxis, as well as blocking LTC_4 -induced bronchoconstriction [23,31-33]. However, in NERD patients, the levels of LXA_4 (downstream product of LXs) and 15-epi-LX in plasma and nasal lavage fluid were reported to decrease after lysine-aspirin nasal challenge [23,31]. A possible reason for the reduced LXA_4 and 15-epi-LX levels in these patients is that the cellular AA resources are already overused for the synthesis of CysLTs [23].

IgE-mediated allergic reactions

NSAIDs can also elicit the production of specific IgE antibodies, which bind to their high-affinity receptors ($Fc\epsilon RI\alpha$ and $Fc\epsilon RII$) on the surface of mast cells and basophils and provide multivalent binding sites for drug antigens. When the drug antigen is cross-linked with IgE bound to the receptors, the mast cells or basophils are stimulated to release preformed mediators (e.g., histamine) and produce new mediators [34]. Although a few studies showed high serum-specific IgE level production by NSAIDs, these results were not replicated and require further investigation [13,35-37].

Despite the conflicting results, one serum-specific IgE that has been confirmed to be involved in NHS is the IgE against thyroid peroxide (TPO), which was first discovered by Altrichter et al. [38] in 2011. More recently, high TPO-specific IgE levels were found in patients with NIUA and NECD compared to healthy controls [39]. Moreover, when TPO was added to basophils isolated from these NIUA and NECD patients, the expression of CD203c (a surface marker for basophil activation) increased [39]. These findings suggest a role of TPO-specific IgE in the pathogenesis of NECD/NIUA via activation of basophils or mast cells [39]. Furthermore, crosslinking of an allergen to the neighboring IgE molecules bound to $Fc\epsilon RI$ can trigger calcium influx, resulting in basophil degranulation [40]. Moreover, in patients with CU and food-dependent, exercise-induced anaphylaxis, aspirin treatment was found to increase phosphorylation of the spleen tyrosine kinase (Syk) in IgE-sensitized basophils [16]. Finally, other NSAIDs (e.g., diclofenac, salicylate, and ketoprofen) were also found to indirectly enhance IgE-mediated histamine release from human basophils through activation of the Syk kinase pathway [40,41].

T cell-mediated mechanisms

NSAIDs can induce delayed-type reactions mediated by T cell activation [2,42]. In the context of maturation signals resulting from drug-related stress, disease, or trauma, dendritic cells (DCs) in the skin and mucous membrane recognize and transport drug antigen complexes to the regional lymph nodes [43]. In the lymph nodes, DCs introduce the drug antigens to naïve T lymphocytes and stimulate the production of antigen-specific T cells. Subsequently, drug antigen-specific T cells migrate to the target organs and secrete cytokines and cytotoxins upon re-exposure to the drugs [43]. Based on the main effector cells involved, T cell-mediated NHS mechanisms are classified into four subtypes: (1) IVa, involving type 1 helper (Th₁) cells and monocytes and the production of interferon γ , interleukin 1 (IL-1), and IL-2 cytokines; (2) IVb, involving Th₂ cells and eosinophils and the production of IL-4, IL-5, and IL-13; (3) IVc, involving cytotoxic T cells and the production of perforin, granzyme B, and Fas ligand; and (4) IVd, involving CD4⁺ T cells, CD8⁺ T cells, and neutrophils, and the production of IL-8 and granulocyte-macrophage colony-stimulating factor [7,42].

Oxidative stress and apoptosis in NHS

Another mechanism underlying NHS involves oxidative stress and the induction of pro-inflammatory mediator production (cytokines and chemokines), which can aggravate airway inflammation, bronchospasm, and mucin secretion [14]. For example, Antczak et al. [44] showed that the levels of 8-isoprostanes, metabolized products from AA that are induced by reactive oxygen species, are increased in the exhaled breath condensate of patients with NERD [45], suggesting a role of oxidative stress in NHS. Moreover, aspirin can trigger apoptosis by inducing oxidative stress as well as by inhibiting expression of the anti-apoptotic protein, Bcl-2 (Bcl2) [17]. NSAIDs may reduce Bcl2 expression by blocking the IL6-IL6R-STAT3 signaling pathway. In turn, decreased Bcl2 expression initiates tumor necrosis factor (TNF)-related apoptosis-inducing ligand and TNF- α generation, which then elicit apoptosis [17].

Platelet activation in NHS

While platelet activation has long been known to be involved in the pathogenic mechanisms of asthma and

urticaria [46,47], its role in NHS has not been studied in depth. However, Palikhe et al. [15] recently found a higher level of soluble P-selectin, which is a marker for activated platelets, in patients with NECD than in those with aspirin-tolerant CU or healthy controls. In addition, while aspirin treatment abolished the expression of soluble P-selectin and P2Y₁₂ on platelets in aspirin-tolerant CU patients, no such inhibitory effect was observed in platelets from NECD patients [15]. These findings suggest that platelet activation contributes to the pathogenesis of NECD.

GENETIC AND EPIGENETIC FACTORS INVOLVED IN NHS

A family history of aspirin intolerance has been reported in 6% of patients, thereby suggesting a role of genetic factors in NHS pathogenesis [48]. However, most genome-wide association and case-control studies have focused on NERD, and only a limited number of studies have investigated genetic factors involved in NECD/NIUA. These studies concentrated on the genes related to the suspected pathomechanisms of the disease, including AA metabolism pathways, IgE, inflammatory mediators, and cytokines, the human leukocyte antigen (HLA) system, and other miscellaneous factors [8,49]. Genetic polymorphisms that have been found to be associated with NHS are shown in Table 2. In addition, epigenetic factors have recently been found to be associated with the pathogenic mechanisms of NHS, which are also summarized in this section.

Genes involved in AA metabolism pathways

Early studies on NHS-associated genes focused on the gene encoding leukotriene C₄ synthase (*LTC₄S*). The C allele of *LTC₄S* -444A>C (rs730012) was reported to be a risk factor for NERD, and was associated with an increase in urine LTE₄ level after aspirin challenge test [50]. In addition, NIUA was found to aggregate in Polish families inheriting the *LTC₄S* -444C allele [51]. However, these findings were not replicated in American, Japanese, or Korean populations [52-54]. Instead, an association of NERD with three single nucleotide polymorphisms (SNPs) in the CysLT receptor 1 gene (*CYSLTR1* -634C>T, -475A>C, and -336A>G) and one SNP

Table 2. Genetic polymorphisms involved in NSAID hypersensitivity

Pathway/gene name	SNPs	Associated clinical phenotype	Population
Arachidonic acid			
<i>LTC4S</i>	-444A>C	C allele was associated with NERD and increased urine LTE ₄ level after aspirin challenge.	Polish [50,51]
<i>CYSLTR1</i>	-634C>T, -475 A>C, -336 A>G	Those SNPs were associated with NERD and could affect gene transcriptional activity.	Korean [55-57]
<i>CYSLTR2</i>	-189 T>C		
<i>COX-2</i>	-765 G>C	This SNP could affect gene transcriptional activity and contribute to PGD ₂ production in NERD patients.	Polish [60]
<i>ALOX5</i>	-1708 G>A, 21 C>T, 270 G>A, 1728 G>A	-1708 A allele was associated with NECD/NIUA.	Korean [53,61]
<i>ALOX15</i>	-272 C>A	Haplotype (GCGA) was associated with NERD. NECD/NIUA	Polish [62]
<i>TBXA2R</i>	-4684 C>T	This SNP affected gene transcriptional activity. C allele was associated with NERD, while TT genotype was associated with NIUA.	Korean [64,65]
	795 T>C	This SNP was associated with NERD and % fall in FEV ₁ after aspirin challenge.	Korean [66]
<i>TBXAS1</i>	141931T>A	This SNP was associated with NERD in a Korean, and NIUA in a Spanish population.	Korean [67] Spain [68]
<i>PTGER2</i>	-616 C>G, -166 G>A	NERD	Korean [65]
<i>PTGER3</i>	-1709 T>A, 173288G>T, 195000A>G	NERD	Korean [65,70]
<i>PTGER4</i>	-1254 A>G	This SNP could affect gene transcriptional activity and was associated with NERD as well as NECD.	Korean [65,69]
<i>PTGIR</i>	1915 T>A	NERD	Korean [65]
Basophil/mast cell and eosinophil activation			
<i>FCER1G</i>	-237 A>G	This SNP was associated with NERD and increased total IgE serum level in NERD patients.	Korean [72]
<i>FCER1A</i>	-344 T>A	This SNP was associated with NECD and increased total IgE serum level in NECD patients. NERD patients carrying CC/CT genotype had a higher serum level of IgE to staphylococcal enterotoxin A than those with CC genotype.	Korean [72,73]
<i>HNMT</i>	939 A>G	This SNP was associated with NECD, could affect HNMT mRNA stability, protein expression, and enzymatic activity of HNMT.	Korean [75]
<i>IL-4</i>	-589 T>C, -33 T>C	These SNPs were associated with NERD and could affect gene transcriptional activity.	Korean [78]
<i>IL-13</i>	-1510 A>C, -1055 C>T	These SNPs were associated with rhinosinusitis in NERD patients.	Korean [79,80]
	110 G>A (Arg 110Gln)	This SNP was associated with increased blood eosinophil count in NERD patients.	Korean [80]
	-1111 C>T	This SNP was associated with NERD.	Japanese [81]
Other inflammatory mediators and cytokines			
<i>IL-5Rα</i>	-5993 G>A	This SNP was associated with the production of serum-specific IgE against staphylococcal enterotoxin A in NERD patients.	Korean [83]
<i>TNF-α</i>	-1031 T>C, -863 C>A	NECD/NIUA	Korean [84]

Table 2. Continued

Pathway/gene name	SNPs	Associated clinical phenotype	Population
	-308 G>A	This SNP was associated with chronic rhinosinusitis and nasal polyposis in NERD.	Hungarian [85]
<i>IL-10</i>	-819 T>C	NERD	Korean [87]
<i>IL-18</i>	-607 A>C -137 G>C	-607 A>C and the haplotype (CG) were associated with NIUA. The (CG) haplotype increased transcriptional activity and neutrophil chemotaxis.	Korean [86]
<i>IL-17A</i>	-737 C>T	NERD	Japan [81]
<i>IL-17Rα</i>	-1075 A>G, -947 A>G, -50 C>T	Major alleles of these SNPs were associated with NERD, higher <i>IL-17RA</i> mRNA and protein expression in CD14 ⁺ peripheral monocytes.	Korean [88]
HLA system			
<i>DPB1*0301</i>	-	This allele was associated with NERD and could interact with TNF-α polymorphisms to increase NERD susceptibility.	Polish [90] Korean [91,92]
<i>DPB1</i>	Rs3128965	This SNP was associated with NERD, increased airway hyper-responsiveness in aspirin challenge test, and increased secretion of 15-HETE from peripheral blood leukocytes.	Korean [92]
	Exm537513	NERD	Korean [94]
<i>DPB1*0401</i>	-	This allele had a protective role against NERD.	Polish [90] German [93]
<i>DQB1*0301, DQB1*011</i>	-	These alleles had a protective role against NERD.	Iran [95]
<i>DQB1*0302, DRB1*04</i>	-	NERD	Iran [95]
<i>DRB1*1302</i>	-	NECD	Korean [96]
<i>DQB1*0609</i> [<i>DRB1*1303/DQB1*0609/DPB1*0201</i>]	-		
<i>Cw4, Cw7</i>	-	These alleles may have a protective role against NECD.	Italian [97]
<i>B44</i>	-	NECD	Italian [97]
Other pathways			
<i>ATF6B</i>	Rs2228628, Rs8111	These SNPs were associated with a decline in %FEV ₁ after aspirin challenge test.	Korean [98]
<i>CYP2C19</i>	681 G>A, 636 G>A	NERD	Japan [99]
<i>DPP10</i>	Rs1704175	This SNP was associated with NERD and the serum level of <i>DPP10</i> in asthmatic patients.	Korean [100]
<i>P2Y12</i>	18 C>T	This SNP was associated with platelet <i>P2Y12</i> expression in NERD patients.	Korean [101]
	742 T>A	This SNP was associated with the nasal secretion level of eosinophil cationic protein in NERD patients.	

NSAID, nonsteroidal anti-inflammatory drug; SNP, single nucleotide polymorphism; LTC₄S, leukotriene C₄ synthase; NERD, NSAID-exacerbated respiratory disease; *LTE₄*, leukotriene E₄; *CYSLTR1*, CysLT receptor 1 gene; *CYSLTR2*, CysLT receptor 2 gene; COX-2, cyclooxygenase 2; *PGD2*, prostaglandin D₂; *ALOX5*, 5-lipoxygenase; NECD, NSAID-exacerbated cutaneous disease; NIUA, NSAID-induced urticaria/angioedema; *TBXA2R*, TXA₂ receptor gene; FEV₁, forced expiratory volume in 1 second; *TBXAS1*, *TBXA1* synthase gene; *PTGER*, prostaglandin E receptor; *PTGIR*, prostaglandin I receptor; *FCER1G*, Fc epsilon receptor 1 gamma; IgE, immunoglobulin E; *FCER1A*, Fc epsilon receptor 1 alpha; HNMT, histamine N-methyltransferase; IL-4, interleukin 4; TNF-α, tumor necrosis factor α; HLA, human leukocyte antigen; 15-HETE, 15-hydroeicosatetraenoic acid; *ATF6B*, activating transcription factor 6β gene; *CYP*, cytochrome P₄₅₀; *DPP10*, dipeptidyl-peptidase 10 gene.

in *CYSLTR2* (-189T>C) was identified in Korean populations. These SNPs were also found to affect gene transcriptional activity [55-57], consistent with the observed increases in *CysLTR1* and *CysLTR2* expression found in nasal mucosal inflammatory cells of NERD patients [58,59].

In Polish subjects with NERD, a promoter SNP in *COX-2* (-765G>C) was found to affect transcriptional activity and contribute to the production of PGD_2 [60], while four SNPs in the 5-LOX gene (*ALOX5* -1708G>A, 21C>T, 270G>A, and 1728G>A) were associated with NERD in a Korean population [53]. The *ALOX5* -1708A allele was also associated with NECD/NIUA in a Korean population [61]. In addition, a promoter SNP in *ALOX15* (-272C>A) was found to be associated with NECD/NIUA in a Spanish population. However, associations of *ALOX12* and *ALOX15* polymorphisms with NHS were not observed in a Korean population [62,63].

The transcriptional function of the -4684C>T SNP in the *TXA2* receptor gene (*TBXA2R*) was reported previously, and was associated with NIUA [64]. Consistently, the *TBXA2R* -4684C allele was shown to be associated with NERD, while the *TBXA2R* -4684TT genotype was associated with NIUA in a Korean population [64,65]. A nonsynonymous SNP of *TBXA2R* (795T>C) was also found to be associated with NERD, as well as with a drop in forced expiratory volume in 1 second (FEV₁) after inhaled aspirin challenge [66]. Similarly, the SNP in intron 9 (rs6962291, 141931T>A) in the *TBXA1* synthase gene (*TBXAS1*) was shown to be associated with NERD in a Korean population [67], as well as with NIUA in a Spanish population [68]. In a Korean population, NERD was significantly associated with genetic polymorphisms of the prostanoid receptors, prostaglandin E receptor 2 (*PTGER2*) (-616C>G, -166G>A), *PTGER3* (-1709T>A), *PTGER4* (-1254A>G), and *PTGIR* (1915T>A) [65]. Moreover, *PTGER4* -1254A>G was found to affect transcriptional activity and was associated with NECD [69]. Another study indicated an association of NERD with rs7543182 (173288G>T) and rs959 (195000A>G) in *PTGER3* [70].

Genes involved in mast cell/basophil and eosinophil activation

The high-affinity IgE receptor (FcεRI) plays a crucial role in the activation and degranulation of mast cells and basophils, resulting in the release of a variety of cytokines

and leukotrienes [71]. Palikhe et al. [72] demonstrated an association of *FCER1G* -237A>G with NERD and increased total IgE serum levels in a Korean population with NERD. On the other hand, the C allele of *FCER1A* -344T>C was associated with NECD and increased total serum IgE level as well as atopy rate in NECD patients [73]. Although no association of *FCER1A* -344T>C with NERD was found, the *FCER1A* -344CT/TT genotype was associated with a higher serum level of IgE specific antibody to Staphylococcal enterotoxin A compared to the CC genotype [72]. Nevertheless, these findings could not be replicated in a recent study with a small population of Taiwanese CU patients [34].

The cellular level of histamine, a key mediator released from basophils/mast cells, is regulated by histamine N-methyltransferase (HNMT) [74]. Therefore, the *HNMT* 939A>G polymorphism may contribute to NECD by influencing *HNMT* mRNA stability, protein expression, and *HNMT* enzymatic activity [75,76]. In addition, *IL-4* and *IL-13* are important cytokines involved in mast cell, basophil, and/or eosinophil activation and function, and share a common receptor subunit (IL-4Rα) [77]. The *IL-4* -589T>C and -33T>C polymorphisms could affect the promoter transcriptional activity, and were shown to be associated with NERD risk [78]. Although associations of *IL-13* polymorphisms (-1510A>C, -1055C>T, and 110G>A [Arg110Gln]) with NERD and NECD were not observed in a Korean population, higher frequencies of rhinosinusitis were noted in NERD patients carrying the *IL-13* -1510A allele, as well as the -1055CC genotype, compared to non-carriers [79,80]. Moreover, the GG genotype of *IL-13* 110G>A was significantly associated with an increased blood eosinophil count in NERD patients [80]. In addition, in a Japanese population, the minor allele of *IL-13* -1111C>T was reported to be associated with NERD [81]. Finally, *IL-5* and its receptor *IL-5R* are necessary for the survival and activation of eosinophils [82], and Losol et al. [83] recently described an association between an *IL-5Rα* polymorphism (-5993G>A) and the production of serum-specific IgE antibodies against Staphylococcal enterotoxin A. However, no association of this *IL-5Rα* polymorphism with NERD susceptibility was found.

Other inflammatory mediators and cytokine-related genes

Among the five SNPs of the *TNF-α* gene investigated in

our previous study ($TNF-\alpha$ -1031T>C, -863C>A, -857C>T, -308G>A, and -238G>A), only two ($TNF-\alpha$ -1031T>C and -863C>A) were significantly associated with NECD/NIUA risk [84]. Associations of chronic rhinosinusitis (CRS) and nasal polyps with $TNF-\alpha$ -308G>A in NERD patients were reported in a Hungarian population, although no association with NERD susceptibility was observed [85]. Kim et al. [86] demonstrated an association of NIUA with $IL-18$ -607A>C, as well as with the haplotype (CG) of $IL-18$ -607A>C and -137G>C, which could increase transcriptional activity and neutrophil chemotaxis. In addition, associations of NERD with other cytokine gene polymorphisms, including $IL-10$ (-819T>C), $IL-17A$ (-737C>T), and $IL-17R\alpha$ (-1075A>G, -947A>G, -50C>T) were also reported in some studies [81,87,88]. These findings suggest that NHS is associated with numerous genetic polymorphisms of inflammatory cytokines, the functions of which in the pathogenesis of NHS should be investigated further.

HLA genes

HLA molecules play a crucial role in T cell activation. Classical HLA class I molecules (HLA-A, HLA-B, and HLA-C) are responsible for presenting drug antigens to CD8⁺ T cells, while HLA class II molecules (HLA-DP, HLA-DQ, and HLA-DR) present antigens to CD4⁺ T cells [89]. Among them, HLA class II alleles are well-known genetic factors associated with NHS. The association of NERD with $HLA-DPB1^*0301$ was first reported in a Polish population, and these findings were then replicated in a Korean population [90,91]. Interestingly, $HLA-DPB1^*0301$ combined with certain $TNF-\alpha$ polymorphisms increases the susceptibility to NERD [91]. More recently, the A allele of rs3128965 in $HLA-DPB1$ was found to be associated with NERD, increased airway hyper-responsiveness after inhaled aspirin challenge test, and increased secretion of 15-HETE from peripheral blood leukocytes treated with AA [92]. In contrast, the $HLA-DPB1^*0401$ allele was reported as a protective factor against NERD susceptibility in studies with Polish and German populations [90,93]. A novel exonic polymorphism (exm537513) in $HLA-DPB1$ associated with NERD was also recently identified [94]. In addition, the combination of exm537513 with six other exonic SNPs from other genes (exm83523, exm1884673, exm538564, exm2264237, exm396794, and exm791954)

provides good predictive value for NERD (area under the curve of 0.75 with 34% sensitivity and 93% specificity) [94]. In addition, Esmailzadeh et al. [95] reported that $HLA-DQB1^*0302$, $HLA-DRB1^*04$, and their haplotype were risk factors of NERD, while $HLA-DQB1^*0301$ and $HLA-DRB1^*011$ were negatively associated with NERD.

NECD has also been found to be associated with $HLA-DRB1^*1302$, $HLA-DQB1^*0609$, and the haplotype of $HLA-DRB1^*1303/DQB1^*0609/DPB1^*0201$ in a Korean population [96]. Furthermore, NECD has also been reported to be associated with HLA class I alleles. Pacor et al. [97] showed that $HLA-Cw4$ and $HLA-Cw7$ could have a protective role against NECD, while $HLA-B44$ was a risk factor. The study by Pacor et al. [97] indicates that HLA class I molecules are involved in the pathogenesis of NHS, and further investigations are required.

Genes related to other pathways

Several genetic risk factors for NERD related to other mechanisms have also been identified. For example, polymorphisms of the activating transcription factor 6 β gene ($ATF6B$ rs2228628 and rs8111) were associated with decline in FEV₁% after inhaled aspirin challenge test [98]. Moreover, two SNPs in cytochrome P450 $CYP2C19$ (681G>A and 636G>A) were found to be associated with NERD and the predicted FEV₁% value [99]. Kim et al. [100] recently reported an association of dipeptidyl-peptidase 10 gene ($DPP10$) rs1704175 (TT genotype) with NERD, and the TT genotype was associated with a higher serum DPP10 level compared to the CC/CT genotypes in asthmatic patients. The purinergic receptor gene polymorphism ($P2RY12$ 742T>C) was associated with the nasal secretion level of an eosinophil cationic protein, while $P2RY12$ 18C>T was associated with platelet P2Y12 expression level in NERD patients [101]. Nevertheless, the underlying mechanisms by which these genes contribute to NERD pathogenesis remain to be elucidated.

Epigenetic mechanisms

A recent study evaluated DNA methylation levels in nasal polyp tissues of patients with NERD and aspirin-tolerant asthma (ATA) [102]. Comparison of the NERD and ATA groups indicated increased methylation levels of 332 loci in 296 genes, while 158 loci in 141 genes had decreased methylation levels. In addition, four genes involved in AA metabolism (PGD synthase [$PGDS$], $PTGES$,

ALOX₅ activating protein [ALOX₅AP], and leukotriene B₄ receptor [LTB₄R]) were found to have different DNA methylation levels between NERD and ATA patients. These findings highlight the value of nasal polyp profiles for suggesting potential susceptibility genes and pathways involved in the pathogenesis of NERD [102].

MANAGEMENT OF PATIENTS WITH NHS

Clinical manifestations

Symptoms of acute NHS include rhinitis/asthma, urticaria/angioedema, and anaphylaxis [82,103]. Patients with NERD have hypersensitivity in the upper (CRS, nasal polyps) and lower (asthma) airways, called Samter's triad [104]. In cases of acute NERD, rhinorrhea and nasal congestion followed by dyspnea and chest tightness appear within 30 to 120 minutes after administration of NSAIDs. The chronic form of NERD, i.e., preexisting inflammation of the upper and lower airways that is exacerbated by ingestion of NSAIDs, usually appears in the third decade of life, and is characterized by a variety of symptoms, including nasal congestion, hyposmia, pansinusitis, laryngospasm, and bronchospasm [105,106]. The phenotype of the disease progresses to severe and corticosteroid-dependent asthma with NHS, and NERD can persist throughout life [107,108]. Cutaneous (such as urticaria) and gastric symptoms may accompany the respiratory symptoms in these patients.

In NECD, symptoms occur within 1 to 4 hours after ingestion of a culprit drug, whereas delayed reactions may occur up to 24 hours. Hives usually disappear within several hours but may persist for days, and some NECD patients may concurrently develop respiratory symptoms [4]. In 30% of CU patients, hypersensitivity reactions occur following ingestion of aspirin or NSAIDs [109]. While aggravation of underlying urticaria by ingestion of the culprit drug can occur in these CU patients, the symptoms may fluctuate depending on how well the underlying disease is controlled [110].

Anaphylaxis is another type of acute reaction in NHS, which is commonly combined with NIUA. Within a few minutes of ingestion or administration of the culprit drug, generalized urticaria and skin swelling may progress into anaphylaxis. Some patients develop both symptoms of NERD and CU [24]. Drugs notorious for

causing anaphylaxis are pyrazolones, ibuprofen, diclofenac, aspirin, and paracetamol [24]. In delayed reactions, symptoms occur within days or even weeks after exposure, but may be induced earlier by reintroduction. Cutaneous reactions (such as fixed drug eruption, severe bullous cutaneous reaction, and maculopapular drug eruption) mostly occur, although rarely organ-specific symptoms (aseptic meningitis, pneumonitis, or nephritis) are reported [24,111].

Diagnosis

The initial step in the diagnosis of NHS is examination of clinical history. Carefully obtained information on symptom descriptions, symptom chronology (including order of incidence or previous exposure to the culprit drugs), and underlying comorbidities (including asthma, CRS, nasal polyps, or CU) are key for NHS diagnosis [6,112]. Adverse reactions to NSAIDs are those where the patient had an adverse drug reaction within 6 hours of NSAID intake, or if such a reaction was recorded by a physician, or if a reaction reoccurred to the same or a distinct NSAID at different times [109]. However, further diagnostic modalities should be applied, as clinical history is never enough to identify the culprit drugs or mechanisms of the reaction. Therefore, skin prick and intradermal tests are useful tools to confirm or exclude sensitization in acute reactions related to an IgE-dependent mechanism, while a patch test is used to identify a delayed reaction related to T cell-mediated responses [6]. However, the sensitivity and specificity of these tests remain unclear and, to date, no standardized tests have been established [6,24]. Moreover, as an *in vitro* study, the basophil activation test may provide useful information; however, it is not used widely due to its low sensitivity and difficulties in setting up the method.

While skin or *in vitro* tests are useful for confirming the culprit drugs, the oral challenge test remains the gold standard for NHS diagnosis [113]. Indeed, an l-lysine-aspirin challenge test and/or aspirin oral challenge test should be performed to confirm the diagnosis of potential NERD patients [106,113]. These oral challenge tests to NSAIDs show very high sensitivity and specificity, exceeding 90% [2]. For the aspirin oral challenge test, the European Academy of Allergy and Clinical Immunology/Global Allergy and Asthma European Network (EAACI/GA²LEN) guidelines outline a 2-day pro-

tolcol where four placebo capsules are administered on the first day, and then four increasing doses of aspirin (71, 117, 312, and 500 mg) are given within a 2-hour interval on the second day, followed by FEV₁ evaluation every 30 minutes [113]. In Korea, the l-lysine-aspirin bronchoprovocation test is widely applied to confirm a diagnosis of NERD; however, to perform these bronchoprovocation tests, the patient should be in a stable condition and the tests should not be performed in a patient with FEV₁ < 70%. Moreover, the bronchoprovocation test should be implemented under the guidance of physicians and technicians equipped with emergency resuscitative equipment [113,114]. The contraindications of challenge tests are acute severe exacerbations, a history of anaphylaxis, or severe cutaneous adverse reactions. It is important to note that although negative results to challenge test may be observed in highly suspicious cases, the diagnosis of NHS should not be excluded, especially in patients on long-term corticosteroid treatment or those with well-controlled underlying diseases.

Recently, Kowalski et al. [2] outlined a simplified, practical diagnostic algorithm for NHS, as follows: (1) the allergist should evaluate if the symptoms are predictable (intolerant) or unpredictable (hypersensitivity); (2) the onset time of the reactions should be assessed; and (3) the pattern of clinical symptoms and underlying chronic disease should be analyzed. In addition, identifying a history of tolerance or intolerance to other NSAIDs helps suggest appropriate NSAIDs for use in challenge tests to confirm or exclude cross-reactivity with other NSAIDs.

Treatment

Immediate NSAID cessation and strict avoidance of the culprit drug and any cross-reacting NSAIDs are of the utmost importance in both early and delayed NHS reactions. Moreover, written information, including lists of culprit and alternative medications, should be provided to NHS patients [7,112].

Most NERD patients have moderate to severe asthma, and medium to high doses of inhaled corticosteroid/long-acting β agonist and leukotriene modifiers should be maintained to control lower respiratory symptoms. The upper airway symptoms related to CRS and/or nasal polyps should be controlled to improve bronchial symptoms, either medically using intranasal corticosteroids,

or surgically if necessary [115]. In cases of severe asthma with frequent asthma exacerbations, biologics, such as anti-IgE antibodies, may have beneficial effects for controlling upper and lower respiratory symptoms [116-118]. The suggested biologics for use in these patients include omalizumab, mepolizumab, and benralizumab [119-121]. For the treatment of CRS/nasal polyps, along with medical treatment (e.g., intranasal corticosteroids), consultation with an ear, nose, and throat specialist for surgical treatment, such as sinus endoscopy, is advised [122].

Systemic steroids and antihistamines may be used during acute, severe cutaneous reactions. However, in severe and delayed reactions (e.g., Stevens-Johnson syndrome [SJS] and toxic epidermal necrolysis [TEN]) withdrawal of the culprit drugs is the only proven treatment to decrease fatality. Although other treatments have been suggested for severe adverse reactions (including systemic corticosteroids, intravenous Igs, or immunosuppressive drugs [e.g., cyclosporin and infliximab]), these treatments remain controversial [24].

Finally, desensitization, which involves the induction of a temporary state of tolerance/unresponsiveness to the drug (mostly aspirin) responsible for NHS, may be an optional treatment for NERD and is expected to improve upper and lower respiratory symptoms [123,124]. Once desensitized, patients should take the drug regularly to maintain tolerance, as intolerance could recur after 2 to 5 days of cessation [124]. Oh et al. [125] reported two cases of NECD that were successfully controlled by aspirin desensitization. Unfortunately, protocols for drug desensitization have not been standardized to date; however, slower protocols tend to be more effective than rush protocols [114]. Despite these successes, the EAA-CI guidelines indicate that desensitization is absolutely contraindicated in cases of severe or threatening delayed drug hypersensitivity reactions (such as SJS, TEN, or drug reactions with eosinophilia and systemic symptoms) [114].

CONCLUSIONS

NHS is a heterogeneous disease with several clinical phenotypes involving various pathogenic mechanisms. In addition to COX pathway alterations, NHS is related

to immune responses (including IgE production and T cell activation), oxidative stress, and platelet activation. While recent genetic studies have identified some predictive markers for NHS susceptibility and treatment response in different populations, these require further investigation. In future, an improved understanding of the functional and genetic/epigenetic pathogenic mechanisms of NHS will help in the development of new diagnostic methods and effective treatments.

Conflict of interest

No potential conflict of interest relevant to this article was reported.

Acknowledgments

This study was supported by a grant from the Korean Health Technology R&D Project, Ministry of Health and Welfare, Republic of Korea (H14C2628).

REFERENCES

- Conaghan PG. A turbulent decade for NSAIDs: update on current concepts of classification, epidemiology, comparative efficacy, and toxicity. *Rheumatol Int* 2012;32:1491-1502.
- Kowalski ML, Makowska JS. Seven steps to the diagnosis of NSAIDs hypersensitivity: how to apply a new classification in real practice? *Allergy Asthma Immunol Res* 2015;7:312-320.
- Kasper L, Sladek K, Duplaga M, et al. Prevalence of asthma with aspirin hypersensitivity in the adult population of Poland. *Allergy* 2003;58:1064-1066.
- Erbagci Z. Multiple NSAID intolerance in chronic idiopathic urticaria is correlated with delayed, pronounced and prolonged autoreactivity. *J Dermatol* 2004;31:376-382.
- Ye YM, Kim MK, Kang HR, et al. Predictors of the severity and serious outcomes of anaphylaxis in Korean adults: a multicenter retrospective case study. *Allergy Asthma Immunol Res* 2015;7:22-29.
- Demoly P, Adkinson NF, Brockow K, et al. International Consensus on drug allergy. *Allergy* 2014;69:420-437.
- Kowalski ML, Makowska JS, Blanca M, et al. Hypersensitivity to nonsteroidal anti-inflammatory drugs (NSAIDs). Classification, diagnosis and management: review of the EAACI/ENDA(##) and GA2LEN/HANNA*. *Allergy* 2011;66:818-829.
- Gomez F, Perkins JR, Garcia-Martin E, Canto G, Cornejo-Garcia JA. Genetic basis of hypersensitivity reactions to nonsteroidal anti-inflammatory drugs. *Curr Opin Allergy Clin Immunol* 2015;15:285-293.
- Dona I, Blanca-Lopez N, Cornejo-Garcia JA, et al. Characteristics of subjects experiencing hypersensitivity to non-steroidal anti-inflammatory drugs: patterns of response. *Clin Exp Allergy* 2011;41:86-95.
- Kowal-Bielecka O, Kowal K, Distler O, Gay S. Mechanisms of disease: leukotrienes and lipoxins in scleroderma lung disease. Insights and potential therapeutic implications. *Nat Clin Pract Rheumatol* 2007;3:43-51.
- Hofmeier KS. Hypersensitivity reactions to modern antiplatelet and anticoagulant drugs. *Allergo J Int* 2015;24:58-66.
- Blanca-Lopez N, Cornejo-Garcia JA, Plaza-Seron MC, et al. Hypersensitivity to nonsteroidal anti-inflammatory drugs in children and adolescents: cross-intolerance reactions. *J Investig Allergol Clin Immunol* 2015;25:259-269.
- Stone SF, Phillips EJ, Wiese MD, Heddle RJ, Brown SG. Immediate-type hypersensitivity drug reactions. *Br J Clin Pharmacol* 2014;78:1-13.
- Cho YS, Moon HB. The role of oxidative stress in the pathogenesis of asthma. *Allergy Asthma Immunol Res* 2010;2:183-187.
- Palikhe S, Palikhe NS, Kim SH, Yoo HS, Shin YS, Park HS. Elevated platelet activation in patients with chronic urticaria: a comparison between aspirin-intolerant and aspirin-tolerant groups. *Ann Allergy Asthma Immunol* 2014;113:276-281.
- Matsuo H, Yokooji T, Morita H, et al. Aspirin augments IgE-mediated histamine release from human peripheral basophils via Syk kinase activation. *Allergol Int* 2013;62:503-511.
- Kacprzak D, Pawliczak R. Does aspirin-induced oxidative stress cause asthma exacerbation? *Arch Med Sci* 2015;11:494-504.
- Bruno A, Tacconelli S, Patrignani P. Variability in the response to non-steroidal anti-inflammatory drugs: mechanisms and perspectives. *Basic Clin Pharmacol Toxicol* 2014;114:56-63.
- Reiss AB, Edelman SD. Recent insights into the role of prostanoids in atherosclerotic vascular disease. *Curr Vasc Pharmacol* 2006;4:395-408.
- Hedi H, Norbert G. 5-Lipoxygenase pathway, dendrit-

- ic cells, and adaptive immunity. *J Biomed Biotechnol* 2004;2004:99-105.
21. Meade EA, Smith WL, DeWitt DL. Differential inhibition of prostaglandin endoperoxide synthase (cyclooxygenase) isozymes by aspirin and other non-steroidal anti-inflammatory drugs. *J Biol Chem* 1993;268:6610-6614.
 22. Vane JR. The mode of action of aspirin and similar compounds. *J Allergy Clin Immunol* 1976;58:691-712.
 23. Kupczyk M, Antczak A, Kuprys-Lipinska I, Kuna P. Lipoxin A4 generation is decreased in aspirin-sensitive patients in lysine-aspirin nasal challenge in vivo model. *Allergy* 2009;64:1746-1752.
 24. Kowalski ML, Asero R, Bavbek S, et al. Classification and practical approach to the diagnosis and management of hypersensitivity to nonsteroidal anti-inflammatory drugs. *Allergy* 2013;68:1219-1232.
 25. Sastre B, del Pozo V. Role of PGE₂ in asthma and non-asthmatic eosinophilic bronchitis. *Mediators Inflamm* 2012;2012:645383.
 26. Ricciotti E, FitzGerald GA. Prostaglandins and inflammation. *Arterioscler Thromb Vasc Biol* 2011;31:986-1000.
 27. Beller TC, Maekawa A, Friend DS, Austen KF, Kanaoka Y. Targeted gene disruption reveals the role of the cysteinyl leukotriene 2 receptor in increased vascular permeability and in bleomycin-induced pulmonary fibrosis in mice. *J Biol Chem* 2004;279:46129-46134.
 28. Paruchuri S, Tashimo H, Feng C, et al. Leukotriene E₄-induced pulmonary inflammation is mediated by the P2Y₁₂ receptor. *J Exp Med* 2009;206:2543-2555.
 29. Gauvreau GM, Parameswaran KN, Watson RM, O'Byrne PM. Inhaled leukotriene E₄, but not leukotriene D₄, increased airway inflammatory cells in subjects with atopic asthma. *Am J Respir Crit Care Med* 2001;164(8 Pt 1):1495-1500.
 30. Cummings HE, Liu T, Feng C, et al. Cutting edge: leukotriene C₄ activates mouse platelets in plasma exclusively through the type 2 cysteinyl leukotriene receptor. *J Immunol* 2013;191:5807-5810.
 31. Sanak M, Levy BD, Clish CB, et al. Aspirin-tolerant asthmatics generate more lipoxins than aspirin-intolerant asthmatics. *Eur Respir J* 2000;16:44-49.
 32. Fierro IM, Colgan SP, Bernasconi G, et al. Lipoxin A₄ and aspirin-triggered 15-epi-lipoxin A₄ inhibit human neutrophil migration: comparisons between synthetic 15 epimers in chemotaxis and transmigration with microvessel endothelial cells and epithelial cells. *J Immunol* 2003;170:2688-2694.
 33. Bonnans C, Levy BD. Lipid mediators as agonists for the resolution of acute lung inflammation and injury. *Am J Respir Cell Mol Biol* 2007;36:201-205.
 34. Hsieh CW, Lee JW, Liao EC, Tsai JJ. A disease marker for aspirin-induced chronic urticaria. *Int J Mol Sci* 2014;15:12591-12603.
 35. Blanca M, Perez E, Garcia JJ, et al. Angioedema and IgE antibodies to aspirin: a case report. *Ann Allergy* 1989;62:295-298.
 36. Bolze S, Bromet N, Gay-Feutry C, Massiere F, Bouliou R, Hulot T. Development of an in vitro screening model for the biosynthesis of acyl glucuronide metabolites and the assessment of their reactivity toward human serum albumin. *Drug Metab Dispos* 2002;30:404-413.
 37. Berkes EA. Anaphylactic and anaphylactoid reactions to aspirin and other NSAIDs. *Clin Rev Allergy Immunol* 2003;24:137-148.
 38. Altrichter S, Peter HJ, Pisarevskaja D, Metz M, Martus P, Maurer M. IgE mediated autoallergy against thyroid peroxidase: a novel pathomechanism of chronic spontaneous urticaria? *PLoS One* 2011;6:e14794.
 39. Shin YS, Suh DH, Yang EM, Ye YM, Park HS. Serum specific IgE to thyroid peroxidase activates basophils in aspirin intolerant urticaria. *J Korean Med Sci* 2015;30:705-709.
 40. Gibbs BF, Rathling A, Zillikens D, Huber M, Haas H. Initial Fc epsilon RI-mediated signal strength plays a key role in regulating basophil signaling and deactivation. *J Allergy Clin Immunol* 2006;118:1060-1067.
 41. Wojnar RJ, Hearn T, Starkweather S. Augmentation of allergic histamine release from human leukocytes by non-steroidal anti-inflammatory-analgesic agents. *J Allergy Clin Immunol* 1980;66:37-45.
 42. Hyman MH. Delayed drug hypersensitivity reactions. *Ann Intern Med* 2004;140:W35.
 43. Gallo PM, Gallucci S. The dendritic cell response to classic, emerging, and homeostatic danger signals. Implications for autoimmunity. *Front Immunol* 2013;4:138.
 44. Antczak A, Montuschi P, Kharitonov S, Gorski P, Barnes PJ. Increased exhaled cysteinyl-leukotrienes and 8-isoprostane in aspirin-induced asthma. *Am J Respir Crit Care Med* 2002;166:301-306.
 45. Dworski R, Murray JJ, Roberts LJ 2nd, et al. Allergen-induced synthesis of F(2)-isoprostanes in atopic asthmatics. Evidence for oxidant stress. *Am J Respir Crit Care Med* 1999;160:1947-1951.

46. Idzko M, Pitchford S, Page C. Role of platelets in allergic airway inflammation. *J Allergy Clin Immunol* 2015;135:1416-1423.
47. Chandrashekar L, Rajappa M, Sundar I, et al. Platelet activation in chronic urticaria and its correlation with disease severity. *Platelets* 2014;25:162-165.
48. Szczeklik A, Nizankowska E, Duplaga M. Natural history of aspirin-induced asthma. AIANE Investigators. European Network on Aspirin-Induced Asthma. *Eur Respir J* 2000;16:432-436.
49. Losol P, Yoo HS, Park HS. Molecular genetic mechanisms of chronic urticaria. *Allergy Asthma Immunol Res* 2014;6:13-21.
50. Sanak M, Pierzchalska M, Bazan-Socha S, Szczeklik A. Enhanced expression of the leukotriene C₄ synthase due to overactive transcription of an allelic variant associated with aspirin-intolerant asthma. *Am J Respir Cell Mol Biol* 2000;23:290-296.
51. Mastalerz L, Setkowicz M, Sanak M, Rybarczyk H, Szczeklik A. Familial aggregation of aspirin-induced urticaria and leukotriene C synthase allelic variant. *Br J Dermatol* 2006;154:256-260.
52. Van Sambeek R, Stevenson DD, Baldasaro M, et al. 5' Flanking region polymorphism of the gene encoding leukotriene C₄ synthase does not correlate with the aspirin-intolerant asthma phenotype in the United States. *J Allergy Clin Immunol* 2000;106(1 Pt 1):72-76.
53. Choi JH, Park HS, Oh HB, et al. Leukotriene-related gene polymorphisms in ASA-intolerant asthma: an association with a haplotype of 5-lipoxygenase. *Hum Genet* 2004;114:337-344.
54. Kawagishi Y, Mita H, Taniguchi M, et al. Leukotriene C₄ synthase promoter polymorphism in Japanese patients with aspirin-induced asthma. *J Allergy Clin Immunol* 2002;109:936-942.
55. Kim SH, Oh JM, Kim YS, et al. Cysteinyl leukotriene receptor 1 promoter polymorphism is associated with aspirin-intolerant asthma in males. *Clin Exp Allergy* 2006;36:433-439.
56. Kim SH, Ye YM, Hur GY, et al. CysLTR₁ promoter polymorphism and requirement for leukotriene receptor antagonist in aspirin-intolerant asthma patients. *Pharmacogenomics* 2007;8:1143-1150.
57. Park JS, Chang HS, Park CS, et al. Association analysis of cysteinyl-leukotriene receptor 2 (CYSLTR₂) polymorphisms with aspirin intolerance in asthmatics. *Pharmacogenet Genomics* 2005;15:483-492.
58. Sousa AR, Parikh A, Scadding G, Corrigan CJ, Lee TH. Leukotriene-receptor expression on nasal mucosal inflammatory cells in aspirin-sensitive rhinosinusitis. *N Engl J Med* 2002;347:1493-1499.
59. Adamusiak AM, Stasikowska-Kanicka O, Lewandowska-Polak A, et al. Expression of arachidonate metabolism enzymes and receptors in nasal polyps of aspirin-hypersensitive asthmatics. *Int Arch Allergy Immunol* 2012;157:354-362.
60. Szczeklik W, Sanak M, Szczeklik A. Functional effects and gender association of COX-2 gene polymorphism G-765C in bronchial asthma. *J Allergy Clin Immunol* 2004;114:248-253.
61. Kim SH, Choi JH, Holloway JW, et al. Leukotriene-related gene polymorphisms in patients with aspirin-intolerant urticaria and aspirin-intolerant asthma: differing contributions of ALOX₅ polymorphism in Korean population. *J Korean Med Sci* 2005;20:926-931.
62. Cornejo-Garcia JA, Jagemann LR, Blanca-Lopez N, et al. Genetic variants of the arachidonic acid pathway in non-steroidal anti-inflammatory drug-induced acute urticaria. *Clin Exp Allergy* 2012;42:1772-1781.
63. Lee HY, Kim SH, Ye YM, Choi GS, Park HS. Lack of association of ALOX₁₂ and ALOX₁₅ polymorphisms with aspirin-exacerbated respiratory disease in Korean patients. *Ann Allergy Asthma Immunol* 2009;103:84-86.
64. Palikhe NS, Kim SH, Lee HY, Kim JH, Ye YM, Park HS. Association of thromboxane A₂ receptor (TBXA₂R) gene polymorphism in patients with aspirin-intolerant acute urticaria. *Clin Exp Allergy* 2011;41:179-185.
65. Kim SH, Kim YK, Park HW, et al. Association between polymorphisms in prostanoid receptor genes and aspirin-intolerant asthma. *Pharmacogenet Genomics* 2007;17:295-304.
66. Kim SH, Choi JH, Park HS, et al. Association of thromboxane A₂ receptor gene polymorphism with the phenotype of acetyl salicylic acid-intolerant asthma. *Clin Exp Allergy* 2005;35:585-590.
67. Oh SH, Kim YH, Park SM, et al. Association analysis of thromboxane A synthase 1 gene polymorphisms with aspirin intolerance in asthmatic patients. *Pharmacogenomics* 2011;12:351-363.
68. Vidal C, Porrás-Hurtado L, Cruz R, et al. Association of thromboxane A₁ synthase (TBXAS₁) gene polymorphism with acute urticaria induced by nonsteroidal anti-inflam-

- matory drugs. *J Allergy Clin Immunol* 2013;132:989-991.
69. Palikhe NS, Sin HJ, Kim SH, et al. Genetic variability of prostaglandin E2 receptor subtype EP4 gene in aspirin-intolerant chronic urticaria. *J Hum Genet* 2012;57:494-499.
 70. Park BL, Park SM, Park JS, et al. Association of PTGER gene family polymorphisms with aspirin intolerant asthma in Korean asthmatics. *BMB Rep* 2010;43:445-449.
 71. Kawakami T, Kashiwakura J, Kawakami Y. Histamine-releasing factor and immunoglobulins in asthma and allergy. *Allergy Asthma Immunol Res* 2014;6:6-12.
 72. Palikhe NS, Kim SH, Cho BY, Ye YM, Hur GY, Park HS. Association of three sets of high-affinity IgE receptor (FcεpsilonR1) polymorphisms with aspirin-intolerant asthma. *Respir Med* 2008;102:1132-1139.
 73. Bae JS, Kim SH, Ye YM, et al. Significant association of FcεpsilonR1α promoter polymorphisms with aspirin-intolerant chronic urticaria. *J Allergy Clin Immunol* 2007;119:449-456.
 74. Yamauchi K, Sekizawa K, Suzuki H, et al. Structure and function of human histamine N-methyltransferase: critical enzyme in histamine metabolism in airway. *Am J Physiol* 1994;267(3 Pt 1):L342-L349.
 75. Kim SH, Kang YM, Kim SH, et al. Histamine N-methyltransferase 939A>G polymorphism affects mRNA stability in patients with acetylsalicylic acid-intolerant chronic urticaria. *Allergy* 2009;64:213-221.
 76. Pope SM, Brandt EB, Mishra A, et al. IL-13 induces eosinophil recruitment into the lung by an IL-5- and eotaxin-dependent mechanism. *J Allergy Clin Immunol* 2001;108:594-601.
 77. McLeod JJ, Baker B, Ryan JJ. Mast cell production and response to IL-4 and IL-13. *Cytokine* 2015;75:57-61.
 78. Kim BS, Park SM, Uhm TG, et al. Effect of single nucleotide polymorphisms within the interleukin-4 promoter on aspirin intolerance in asthmatics and interleukin-4 promoter activity. *Pharmacogenet Genomics* 2010;20:748-758.
 79. Palikhe NS, Kim SH, Choi GS, Ye YM, Park HS. No evidence of association between interleukin-13 gene polymorphism in aspirin intolerant chronic urticaria. *Allergy Asthma Immunol Res* 2009;1:36-40.
 80. Palikhe NS, Kim SH, Cho BY, et al. IL-13 gene polymorphisms are associated with rhinosinusitis and eosinophilic inflammation in aspirin intolerant asthma. *Allergy Asthma Immunol Res* 2010;2:134-140.
 81. Kohyama K, Abe S, Kodaira K, et al. IL-13 and IL-17A gene polymorphisms in Japanese patients with aspirin-exacerbated respiratory disease. *Ann Allergy Asthma Immunol* 2011;107:510-516.
 82. Choi JH, Kim MA, Park HS. An update on the pathogenesis of the upper airways in aspirin-exacerbated respiratory disease. *Curr Opin Allergy Clin Immunol* 2014;14:1-6.
 83. Losol P, Kim SH, Shin YS, Ye YM, Park HS. A genetic effect of IL-5 receptor α polymorphism in patients with aspirin-exacerbated respiratory disease. *Exp Mol Med* 2013;45:e14.
 84. Choi JH, Kim SH, Cho BY, et al. Association of TNF-alpha promoter polymorphisms with aspirin-induced urticaria. *J Clin Pharm Ther* 2009;34:231-238.
 85. Szabo K, Kiricsi A, Revesz M, et al. The -308 G>A SNP of TNFA is a factor predisposing to chronic rhinosinusitis associated with nasal polyposis in aspirin-sensitive Hungarian individuals: conclusions of a genetic study with multiple stratifications. *Int Immunol* 2013;25:383-388.
 86. Kim SH, Son JK, Yang EM, Kim JE, Park HS. A functional promoter polymorphism of the human IL18 gene is associated with aspirin-induced urticaria. *Br J Dermatol* 2011;165:976-984.
 87. Palikhe NS, Kim SH, Jin HJ, Nam YH, Park HS. Association of interleukin 10 promoter polymorphism at -819 T>C with aspirin-induced urticaria in a Korean population. *Ann Allergy Asthma Immunol* 2011;107:544-546.
 88. Park JS, Park BL, Kim MO, et al. Association of single nucleotide polymorphisms on Interleukin 17 receptor A (IL17RA) gene with aspirin hypersensitivity in asthmatics. *Hum Immunol* 2013;74:598-606.
 89. Bronietzki AW, Schuster M, Schmitz I. Autophagy in T-cell development, activation and differentiation. *Immunol Cell Biol* 2015;93:25-34.
 90. Dekker JW, Nizankowska E, Schmitz-Schumann M, et al. Aspirin-induced asthma and HLA-DRB1 and HLA-DPB1 genotypes. *Clin Exp Allergy* 1997;27:574-577.
 91. Choi JH, Lee KW, Oh HB, et al. HLA association in aspirin-intolerant asthma: DPB1*0301 as a strong marker in a Korean population. *J Allergy Clin Immunol* 2004;113:562-564.
 92. Kim SH, Ye YM, Lee SK, et al. Association of TNF-alpha genetic polymorphism with HLA DPB1*0301. *Clin Exp Allergy* 2006;36:1247-1253.
 93. Lympany PA, Welsh KI, Christie PE, Schmitz-Schumann M, Kemeny DM, Lee TH. An analysis with sequence-spe-

- cific oligonucleotide probes of the association between aspirin-induced asthma and antigens of the HLA system. *J Allergy Clin Immunol* 1993;92(1 Pt 1):114-123.
94. Shin SW, Park BL, Chang H, et al. Exonic variants associated with development of aspirin exacerbated respiratory diseases. *PLoS One* 2014;9:e111887.
 95. Esmaeilzadeh H, Nabavi M, Amirzargar AA, et al. HLA-DRB and HLA-DQ genetic variability in patients with aspirin-exacerbated respiratory disease. *Am J Rhinol Allergy* 2015;29:e63-e69.
 96. Kim SH, Choi JH, Lee KW, et al. The human leucocyte antigen-DRB1*1302-DQB1*0609-DPB1*0201 haplotype may be a strong genetic marker for aspirin-induced urticaria. *Clin Exp Allergy* 2005;35:339-344.
 97. Pacor ML, Di Lorenzo G, Mansueto P, et al. Relationship between human leucocyte antigen class I and class II and chronic idiopathic urticarial associated with aspirin and/or NSAIDs hypersensitivity. *Mediators Inflamm* 2006;2006:62489.
 98. Park TJ, Kim JH, Pasaje CF, et al. Polymorphisms of AT-F6B are potentially associated with fev1 decline by aspirin provocation in asthmatics. *Allergy Asthma Immunol Res* 2014;6:142-148.
 99. Kohyama K, Abe S, Kodaira K, et al. Polymorphisms of the CYP2C19 gene in Japanese patients with aspirin-exacerbated respiratory disease. *J Allergy Clin Immunol* 2011;128:1117-1120.
 100. Kim SH, Choi H, Yoon MG, Ye YM, Park HS. Dipeptidyl-peptidase 10 as a genetic biomarker for the aspirin-exacerbated respiratory disease phenotype. *Ann Allergy Asthma Immunol* 2015;114:208-213.
 101. Losol P, Palikhe NS, Lee JW, et al. Association of P2RY12 polymorphisms with eosinophil and platelet activation in patients with aspirin-exacerbated respiratory disease. *Ann Allergy Asthma Immunol* 2015;114:423-424.
 102. Cheong HS, Park SM, Kim MO, et al. Genome-wide methylation profile of nasal polyps: relation to aspirin hypersensitivity in asthmatics. *Allergy* 2011;66:637-644.
 103. Choi JH, Kim JH, Park HS. Upper airways in aspirin-exacerbated respiratory disease. *Curr Opin Allergy Clin Immunol* 2015;15:21-26.
 104. Samter M, Beers RF Jr. Concerning the nature of intolerance to aspirin. *J Allergy* 1967;40:281-293.
 105. Graefe H, Roebke C, Schafer D, Meyer JE. Aspirin sensitivity and chronic rhinosinusitis with polyps: a fatal combination. *J Allergy (Cairo)* 2012;2012:817910.
 106. Lee RU, Stevenson DD. Aspirin-exacerbated respiratory disease: evaluation and management. *Allergy Asthma Immunol Res* 2011;3:3-10.
 107. Jenkins C, Costello J, Hodge L. Systematic review of prevalence of aspirin induced asthma and its implications for clinical practice. *BMJ* 2004;328:434.
 108. Baenkler HW. Salicylate intolerance: pathophysiology, clinical spectrum, diagnosis and treatment. *Dtsch Arztebl Int* 2008;105:137-142.
 109. Asero R, Bavbek S, Blanca M, et al. Clinical management of patients with a history of urticaria/angioedema induced by multiple NSAIDs: an expert panel review. *Int Arch Allergy Immunol* 2013;160:126-133.
 110. Parejo PA, Garcia-Agundez JA, Cornejo-Garcia JA, et al. Association study of functional polymorphisms in genes involved in histamine homeostasis and multiple NSAID: triggered urticaria and/or angioedema and anaphylaxis in patients without pre-existing chronic urticaria (MNSAID-UA). *J Allergy Clin Immunol* 2013;131(2 Suppl):AB169.
 111. Johansson SG, Bieber T, Dahl R, et al. Revised nomenclature for allergy for global use: report of the Nomenclature Review Committee of the World Allergy Organization, October 2003. *J Allergy Clin Immunol* 2004;113:832-836.
 112. Simons FE, Arduzzo LR, Biló MB, et al. 2012 Update: World Allergy Organization Guidelines for the assessment and management of anaphylaxis. *Curr Opin Allergy Clin Immunol* 2012;12:389-399.
 113. Nizankowska-Mogilnicka E, Bochenek G, Mastalerz L, et al. EAACI/GA2LEN guideline: aspirin provocation tests for diagnosis of aspirin hypersensitivity. *Allergy* 2007;62:1111-1118.
 114. Scherer K, Brockow K, Aberer W, et al. Desensitization in delayed drug hypersensitivity reactions: an EAACI position paper of the Drug Allergy Interest Group. *Allergy* 2013;68:844-852.
 115. Mullol J, Picado C. Rhinosinusitis and nasal polyps in aspirin-exacerbated respiratory disease. *Immunol Allergy Clin North Am* 2013;33:163-176.
 116. Pinto JM, Mehta N, DiTineo M, Wang J, Baroody FM, Naclerio RM. A randomized, double-blind, placebo-controlled trial of anti-IgE for chronic rhinosinusitis. *Rhinology* 2010;48:318-324.
 117. Vennera Mdel C, Picado C, Mullol J, Alobid I, Bernal-Sprekelsen M. Efficacy of omalizumab in the treatment of nasal polyps. *Thorax* 2011;66:824-825.

118. Mesonjesi E, Ziu EP, Priftanji A, Qama D. Omalizumab for severe allergic asthma: a life changing drug? *Clin Transl Allergy* 2015;5(Suppl 2):P18.
119. Brightling CE, Chanaz P, Leigh R, et al. Efficacy and safety of tralokinumab in patients with severe uncontrolled asthma: a randomised, double-blind, placebo-controlled, phase 2b trial. *Lancet Respir Med* 2015;3:692-701.
120. Darveaux J, Busse WW. Biologics in asthma: the next step toward personalized treatment. *J Allergy Clin Immunol Pract* 2015;3:152-160.
121. Fainardi V, Pisi G, Chetta A. Mepolizumab in the treatment of severe eosinophilic asthma. *Immunotherapy* 2016;8:27-34.
122. Riechelmann H; Europäischen Akademie für Allergie und Klinische Immunologie (EAACI) und der European Rhinologic Society (ERS). Chronic rhinosinusitis: EPOS 2012 part I. *Laryngorhinootologie* 2013;92:193-201.
123. Cernadas JR, Brockow K, Romano A, et al. General considerations on rapid desensitization for drug hypersensitivity: a consensus statement. *Allergy* 2010;65:1357-1366.
124. White AA, Stevenson DD. Aspirin-exacerbated respiratory disease: update on pathogenesis and desensitization. *Semin Respir Crit Care Med* 2012;33:588-594.
125. Oh MS, Seong GM, Lee HS, Lim GC, Lee J. Two cases of aspirin sensitive, intractable chronic urticaria successfully controlled by aspirin desensitization. *Korean J Asthma Allergy Clin Immunol* 2012;32:51-55.